# Computational Biology DNA Sequencing

Department of Mathematics Stockholm University

## Copying DNA:

Polymerase chain reaction (PCR)

- Sequencing DNA:
  - Sanger Sequencing [AKA 1st generation sequencing]
  - Next/2nd-generation sequencing (NGS) [AKA Massive parallel sequencing]
  - 3rd-generation [AKA long-read sequencing]

#### **Polymerase chain reaction (PCR)**

- used to copy DNA ►
- Invented by Kary Mullis (Nobel prize 1993)
- Input: a DNA "template" t to copy, primers, polymerase, bases A, C, G, T,

Process: *n* "cycles" (see right)

Output: roughly  $2^n$  copies of t

Per cycle there are 3 phases:



Denaturate: 94-98 °C for 20-30 s

Anneal: 50-65 °C for 20-40 s

Extension: 75-80 °C

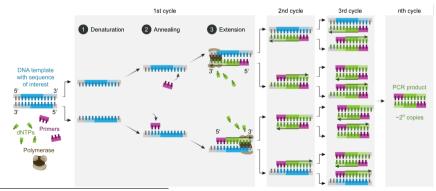


fig taken from https://en.wikipedia.org/wiki/Polymerase\_chain\_reaction

#### **Sanger Sequencing**

- used to read "small ( 500bp)" DNA sequences
- Invented by Fredrick Sanger and coworkers, 1977 (Nobel prize 1980)
- Input: copies of DNA split into 4 test tubes that contains primers, polmerase, bases, "modified bases A, C, T, G"

Each tube contains all bases and ONE "modified base"  $I \in \{A, C, G, T\}$ 

Process (Basic Idea): "modified base" *I* ensures that when added during reading process of one DNA-copy, the reading process stops.

Having multiple copies and the four tubes, this ensures, that (with high probability) the tupe *I* contains all single strands that end with *I*.

gel electrophoresis: reads are negative charged and small reads get "closer" to positive pol (proportional to their length)

Output: the read of the input DNA

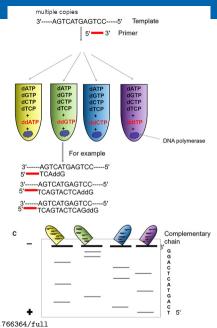
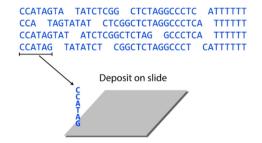


fig taken from https://www.frontiersin.org/articles/10.3389/fmicb.2021.766364/full

#### Next-generation sequencing (NGS)

- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

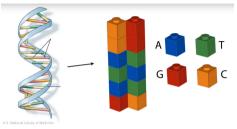


figs taken from Ben Langmead "ADS1: Sequencing by Synthesis" (youtube)

#### Next-generation sequencing (NGS)

- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..



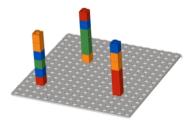
Double stranded DNA (double helix) Double stranded DNA (lego version)

#### figs taken from Ben Langmead "ADS1: Sequencing by Synthesis" (youtube)

#### Next-generation sequencing (NGS)

- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

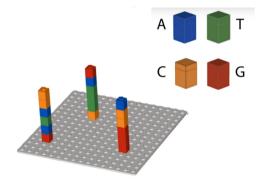


figs taken from Ben Langmead "ADS1: Sequencing by Synthesis" (youtube)

#### Next-generation sequencing (NGS)

- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

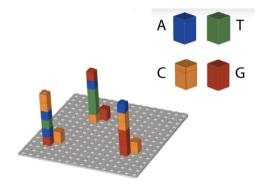


#### Next-generation sequencing (NGS)

- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

Process (Basic Idea): when bases with terminators bind, no further base can be added.



figs taken from Ben Langmead "ADS1: Sequencing by Synthesis" (youtube)

### Next-generation sequencing (NGS)

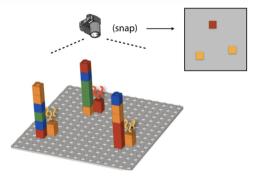
- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

Process (Basic Idea): when bases with terminators bind, no further base can be added.

terminators are engineered to glow a particular color (A, C, G, T)

 $\rightarrow$  take photo



figs taken from Ben Langmead "ADS1: Sequencing by Synthesis" (youtube)

#### Next-generation sequencing (NGS)

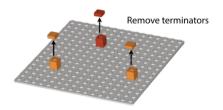
- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

Process (Basic Idea): when bases with terminators bind, no further base can be added.

```
terminators are engineered to glow a particular color (A, C, G, T)
```

 $\rightarrow$  take photo



### Next-generation sequencing (NGS)

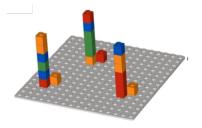
- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

Process (Basic Idea): when bases with terminators bind, no further base can be added.

```
terminators are engineered to glow a particular color (A, C, G, T)
```

 $\rightarrow$  take photo



## Next-generation sequencing (NGS)

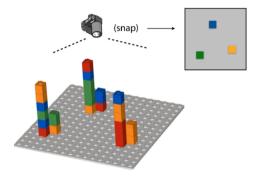
- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

Process (Basic Idea): when bases with terminators bind, no further base can be added.

```
terminators are engineered to glow a particular color (A, C, G, T)
```

 $\rightarrow$  take photo



figs taken from Ben Langmead "ADS1: Sequencing by Synthesis" (youtube)

### Next-generation sequencing (NGS)

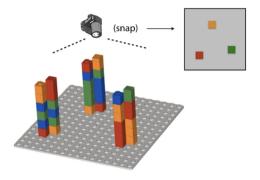
- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

Process (Basic Idea): when bases with terminators bind, no further base can be added.

```
terminators are engineered to glow a particular color (A, C, G, T)
```

 $\rightarrow$  take photo



## Next-generation sequencing (NGS)

- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

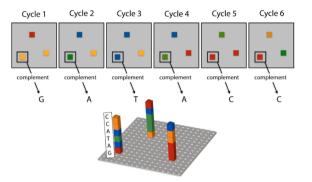
Process (Basic Idea): when bases with terminators bind, no further base can be added.

```
terminators are engineered to glow a particular color (A, C, G, T)
```

 $\rightarrow$  take photo

after taking photo, terminators are removed and process is repeated.

Output: the read of the **multiple** input DNAs (photos of each cycle)



## Next-generation sequencing (NGS)

- used to read multiple "small ( 500bp)" DNA sequences
- Several methods exits, one is the "Illumina sequencing process":

Input: copies of **multiple** DNA (fragments) placed on a slide, bases, terminators, polymerase, ..

Process (Basic Idea): when bases with terminators bind, no further base can be added.

terminators are engineered to glow a particular color (A, C, G, T)

 $\rightarrow$  take photo

after taking photo, terminators are removed and process is repeated.

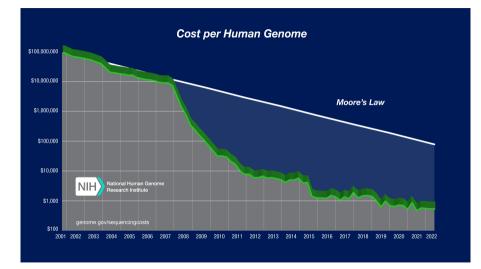
Output: the read of the **multiple** input DNAs (photos of each cycle)

#### Key feature:

massively parallel, photograph captures all templates simultaneously (billions of DNA templates on a slide)

fig taken from https://theory.labster.com/ngs-experiment/

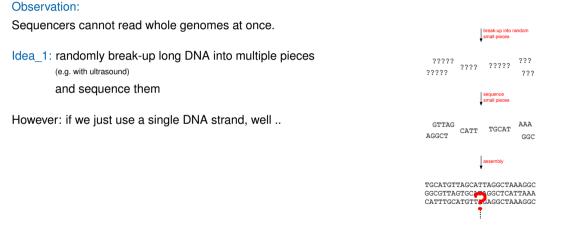




- Copying DNA:
  - Polymerase chain reaction (PCR)
- Sequencing DNA:
  - Sanger Sequencing [AKA 1st generation sequencing]
  - Next/2nd-generation sequencing (NGS) [AKA Massive parallel sequencing]
  - 3rd-generation [AKA long-read sequencing] (currently under active development\*, can read more than 10000 bp)

To recall, humanDNA  $3.2 \times 10^9$  bp, Carsonella ruddii DNA 159 662 bp Observation: Whole genomes cannot be read at once.

<sup>\*</sup>Marx, V. Method of the year: long-read sequencing. Nat Methods 20, 6–11 (2023).



#### Observation:

Sequencers cannot read whole genomes at once.

Idea\_1: randomly break-up long DNA into multiple pieces (e.g. with ultrasound) and sequence them

However: if we just use a single DNA strand, well ..

Idea\_2: Produce multiple copies of DNA first and then apply Idea\_1

- $\implies$  results in overlapping reads
- $\implies$  assempty (here smart computational methods are needed!)

unknown DNA PCR break-up into random small nieces ????? ????? 2222 >>>>> ???? ??????? 222 ??? ?????????? sequence small pieces GTTAG TGCAT CATT AGGCT CATC TTAGCAT TAGGCTAA ATG TGC GCTAAAGGC assembly TGC TGCAT CATG ATG GTTAG TTAGCAT GCATT TAGGCTAA AGGCT GCTAAAGGC

Seguence Assembly

For a given set  $\zeta = \{S_1, \dots, S_N\}$  of strings (=reads of fragments of DNA *D*), a superstring is a string *S* that contains all  $S_i$  as substrings.

Trivially, we could concatenate all strings in  $\zeta$  to get superstring *S*. However, having say  $\sim 10^6$  copies of DNA *D* fragmented and sequenced, we get then a string *S* of length  $|S| \sim |D| \times 10^6$   $\implies$  far away from *D*.

In the assembly problem, we want to find a superstring that "best represents" D.

There are several ways on how to define "best represents" !!

We start with considering following problem:

# Shortest Common Superstring Problem (SCS):

For a given  $\zeta = \{S_1, \dots, S_N\}$  find a superstring *S* of shortest length.

SCS is NP-hard. So we focus ways to approximate solutions

 $\implies$  overlap graphs and Greedy\_SCS (board)

$$\frac{Basics}{Basics} : = tring S = x_{1}... - x_{m}, |S| = m$$

$$S[i..j] = x_i x_{i+1} - x_j'$$

$$S[i..j] = x_i x_{i+1} - x_j'$$

$$S(i) = x_i$$

$$S(i) = x_i$$

$$S[i..j] = prefix of S ending at j$$

$$S[j..m] = Suffix of S stating at j$$

-> shortest supershing S is

.

Theorem (Gallant dal 1980): SCS is NP-hard => no hope for polynomial-time exact algorithm (P=NP) Q: Can ue approximate a solution? In what follows: J= {S1... SN } "Subshing-free" of Siasj & Siffsj Viaj. [IF S := S; or S : E S; , we can remove S; in preprocess step] has no impact on SCS! we will see later, this can be done in O(N·11511) time 11511 = E ISI via Suffix-ses frees

DEF:

For all Si, Sj 
$$\in \int = \frac{1}{2}S_{1}...S_{NS}^{2}$$
 there is  
a longest substring Vij of Vij is suffix of Si  
 $\downarrow$  prefix of Sj  
 $\downarrow$  that is  $S_{i}^{-} = \sqcup V_{ij}$   
 $S_{j}^{-} = V_{ij}^{-} W$  ( $v_{ij}^{-} = \varepsilon = empty shing$   
 $i^{2} possible !$ )  
 $V_{ij}^{-} called orulap of S; & Sj
 $ov(i,j) = |v_{ij}|$  ( $i \quad V_{ij}^{-} \neq V_{j}^{-} c possible !$ )  
 $p(i,j) = |u_{ij}|$  ( $i \quad V_{ij}^{-} \neq V_{j}^{-} c possible !$ )  
 $p(i,j) = |u_{ij}|$   
 $merge(i,j) = u_{ij}^{-} w$   
 $pref(i,j) = u_{ij}^{-} w$$ 

EXMPL  

$$S_{n} = AT AT$$
,  $S_{2} = ATTT$   
 $V_{n2} = AT$   
 $vv (A_{1}2) = 2$   
 $p (A_{2}2) = 2$   
 $S_{2} = ATTT$   
 $S_{n} = ATAT$   
 $V_{2n} = E (compty)$ 

or (2.1) = 0  

$$P(2,1) = 4$$
  
 $Ef_2 = ATTT = UV_{21}$   
 $Marge(2,1) = ATTTATAT$ 

$$\frac{DEF}{S} = \operatorname{graph} G_{g} = (V_{i} E) \text{ or } \mathcal{U} = \{S_{n}, S_{i} \vee S\}:$$

$$V = \{S_{i} \in \mathbb{C} \setminus \{S_{i} \setminus S\} \in S\} = \{S_{i} \setminus S\} \setminus \{S_{i} \setminus S\}: S \in S\}$$

$$"no loops"$$

$$"prehr graph" : arc S_{i} \longrightarrow S_{j} gete weight p(n; j)$$

$$"or (no graph" : unime units of (no used no units up or (n; j)).$$

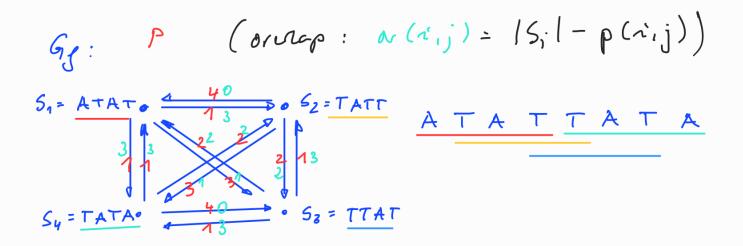
$$I can be constructed via O(no used up to used up to$$

 $E \times MPL$ : (1)  $f = \{ S_1 = A \top A \top, S_2 = A \top T \top f \}$ 

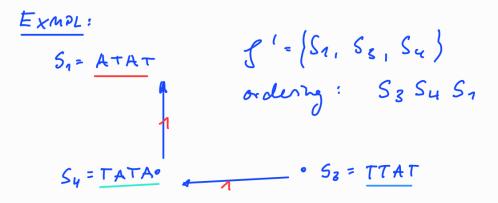
$$G_{g}: \begin{array}{c} \frac{40}{S_{1} + 2} \\ S_{1} + 2 \\ S_{2} \end{array} = \begin{array}{c} w_{ij} \\ p(ij) \\ p(ij) \\ s_{1} + 1 \\ s_{2} + 2 \\ s_{2} \end{array} = \begin{array}{c} w_{ij} \\ p(ij) \\ s_{1} + 1 \\ s_{2} + 1 \\ s_{3} + 2 \\ s_{4} \end{array} = \begin{array}{c} w_{ij} \\ p(ij) \\ s_{1} + 1 \\ s_{2} + 1 \\ s_{3} + 2 \\ s_{4} + 2 \\ s_{5} + 2 \\ s$$

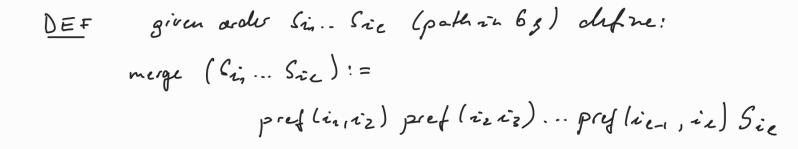
Observe min-length superstring 
$$S$$
 for  $S_1, S_2$   
is  
 $S = ATATIT = \frac{2}{S_1} S_2$   
is  $S_1 = S_2$   
is are where  
perfix-length min./  
derelep mex.

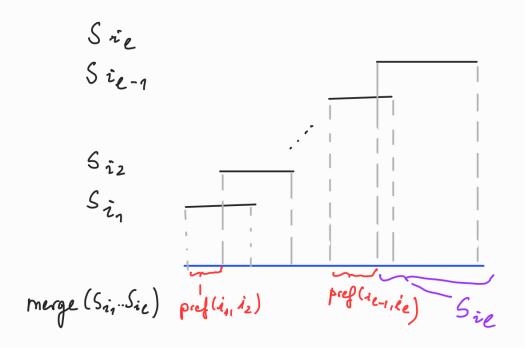
(2)  $\int_{\mathcal{A}} \{ ATAT , TATA, TATT , TTAT \}$ 



1:1 correspondence betreen orderings of elements in S'ES & paths in 6g.







 $EXMPL(above) \qquad order S_2, S_4, S_1$   $S_1 = \underbrace{T A T A}_{S_4} = \underbrace{A T A T}_{S_3} = \underbrace{T T A T}_{S_3} = \underbrace{T T A T}_{S_3}$   $merge (S_2 S_4 S_1) = \underbrace{T T A T A}_{S_1} = \underbrace{S_1}_{S_1}$   $pref (3.2) \mid S_1$  pref (4,1)

By def: 
$$[merge(Sin-Sie)] = \sum_{i=1}^{l} p(\overline{v}, \overline{v}+1) + |S_{\overline{v}e}|$$

EXMPL

$$J = \left\{ \begin{array}{ccc} ATAT & TATA \\ Gg : & Ar(n',j) \end{array} \right\}$$

$$S_{1} = ATAT \circ \begin{array}{c} a & 0 \\ \hline & 3 \\ \hline \hline & 3 \\ \hline & 3 \\ \hline \\$$

max or where 
$$i \in 3$$
 eq  $(S_{i1}S_{i1})$ ,  $(S_{i1}G_{i1})$ ,  $(S_{2}S_{i1})$ ,  $(S_{3}S_{2})$ .  
Aalm one of Mum & merge.  
Choose here  $(S_{7}, S_{2})$ :  $S_{i} \neq S_{2}$   
=?  $\int g = \{ATATT, TTAT, TATA\}$   
merge $(S_{1}S_{2})$   $S_{2}$   $S_{1}$   
 $\int g' = ATATT$   
 $S_{i} = TATA^{*}$   
 $S_{i} = TATATT$   
 $S_{i} = TATATT$   
 $S_{i} = TATATT$   
 $S_{i} = TATATT$   
 $S_{i} = TATA^{*}$   
 $S_{i} = TTAT^{*}$   
 $S_{i} = TATA^{*}$   
 $S_{i} = TTAT^{*}$   
 $S_{i} = TATA^{*}$   
 $S_{i} = TTAT^{*}$   
 $S_{i} = TTAT^{*}$   

Assame now, for conhadiction, that after  
some step 
$$N \ge 1$$
, 5 does not redisty  
Condition in the beams.  
Put  $J_{N-1} = S$  after applying N-1 steps of greeds.  
By assumption in  $J_{N-1}$  everything ok!  
Wiley let  $S', S'' \in J_{N-1}$  all or  $(S', S'') = \max$  and and a power  
all elements in  $S$   
l greedy chooses  $S' l S''$  to be marged.  
 $= S$   
 $J_N = J_{N-1} \setminus \{S', S''\} \cup \{\max (S', S'')\}$   
 $= S$ 

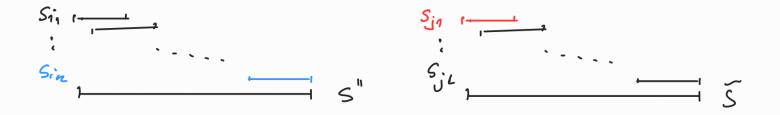
Situation is as follows:

$$S = merge(S', S'')$$
  

$$S'' = merge(S_{i_1}, S_{i_k}) \quad k \ge 1, \quad S_{i_r} \in S_0$$
  

$$S'' = merge(S_{j_n}, S_{i_k}) \quad k \ge 1, \quad S_{j_r} \in S_0$$

& since condition in lene violeted we have ou(S,S) = ou(Sik,Sjn) Landez arguments for course or (Sis) = ...]

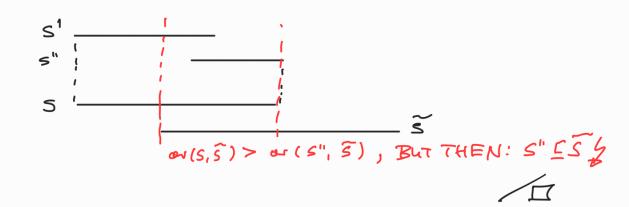


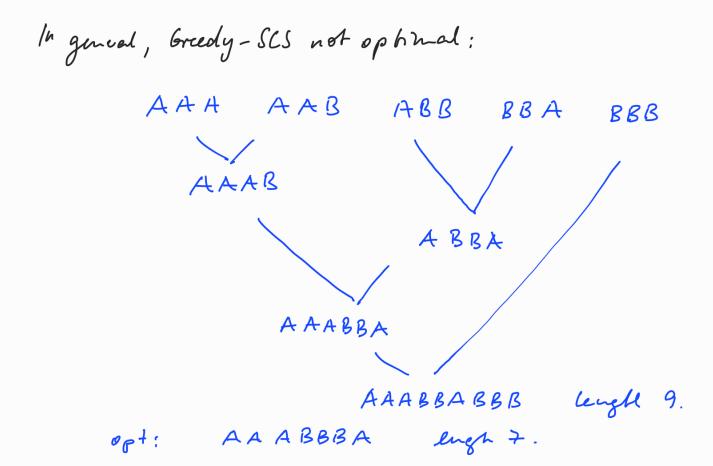
By definition :  $S_{ik} \sqsubseteq S = merge (S', S'')$ de  $S_{ij} \sqsubseteq S$ 

> ->  $ov(S, \tilde{S}) \ge ov(Sik, Sjn)$ A mue  $n \neq n$

By Jud-ensumption (all fine in 
$$S_{N-1} \& \bar{S}, S'' \in S_{N-1}$$
)  
we have:  
 $or(S'', \bar{S}) = or(Si_{K}, S_{j_{n}}) \ll or(S_{j}, \bar{S})$ 

=> must look like:





ie, never worke than 4 times opt-solution. Ewill out proof ]

One particular problem in practice : represts.

In genomes ofter repeated regions.

Genomes often consist of repeated regions!

**Example:** Here,  $\zeta$  = set of all substrings of size 6.

```
Greedy SCS on 6-mers of a long long long time
```

```
ng_lon _long_ a_long long_l ong_ti ong_lo long_t g_long g_time ng_tim
ng_time ng_lon _long_ a_long long_l ong_ti ong_lo long_t g_long
ng_time g_long_ ng_lon a_long long_l ong_lo long_t
ng_time long_ti g_long_ ng_lon a_long long_l ong_lo
ng_time ong_lon long_ti g_long_ a_long long_l
ong_lon long_time g_long_ a_long
long_lon long_time g_long_ a_long
long_lon g_long_time a_long
long_long_time a_long
a_long_long_time a_long
```

The final superstring is shorter than the original "genome"

```
Genomes often consist of repeated regions!
                                                                       & 131=16 for all examples
Example: Here, \zeta = set of all substrings of size 6.
                                        a_long_long_time
                                                                 a_long_long_long_time
                                          long long ta_long long_long_long_timelong long tilong long_log_timelong log_tilong_ong_log_timelong l ng_timlong_l ng_lonlong l g_timeong_log_longong_long_lon_long
                                        a_long long_t
                                            ong_lo
                                                                       g_long long_t
                                             ng_lon
ng_lon
g_long
                                                                         _long_ ong_ti
                                              g_long
                                               _long_
_long_
                                             a_long_long_long_long_time
                                            ong_ti
```

To work with such problems one may employ: DeBruijn-graphs and Eulerian Paths. (board)

$$\frac{E + hPL}{J} = \{AAAA, AABB, BBA, ABBBS \}$$

$$\rightarrow J = \{AAAA, AABB, BBA, ABBBS \}$$

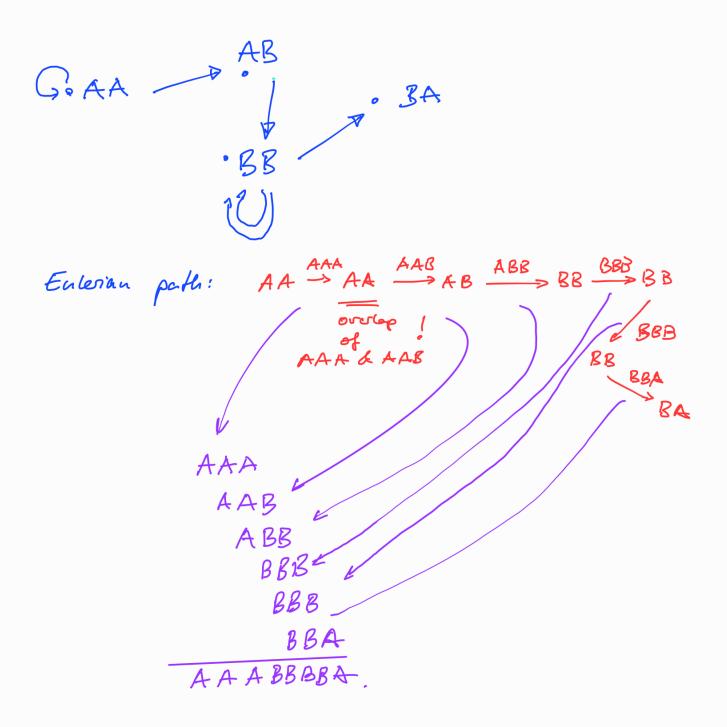
$$\rightarrow J_{3} = \{AAA, AAB, ABB, BBB, BBB, BBA3$$

Subdivide each k-mer samse into left k-1 mer samsker right k-2-mer szinsker fræse become the vertices an Oebruijn grogsk Gre arc (V, W) in Gre if V is L-k-iner w is R-k-1 ner of some kmer ar Jr.

EthPL: (unknown) DNA	AAABBBBA
all 3mers; Jz=	(AAA, AAB, ABB, BBB, BBB, BBA]
can eg be obtained from reads of frogments.	
3-mas AAA -	> L K-Imer AA R k-Imer AA
A A B	L - u - AA R - u - AB
ABB	L AB R BB
(2×) BBB	L BB R BB//
BBA	L BB / R BA /

Verices on gz: GOAA AB BB BA 2) edges (rimplichely) represent 3-mos AB ABB BB

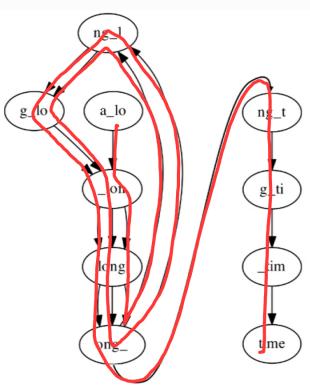
Enterian path in a directed graft is a path that "visits" each edge exactly once.



De Bruijn graph (*k*=5) for:

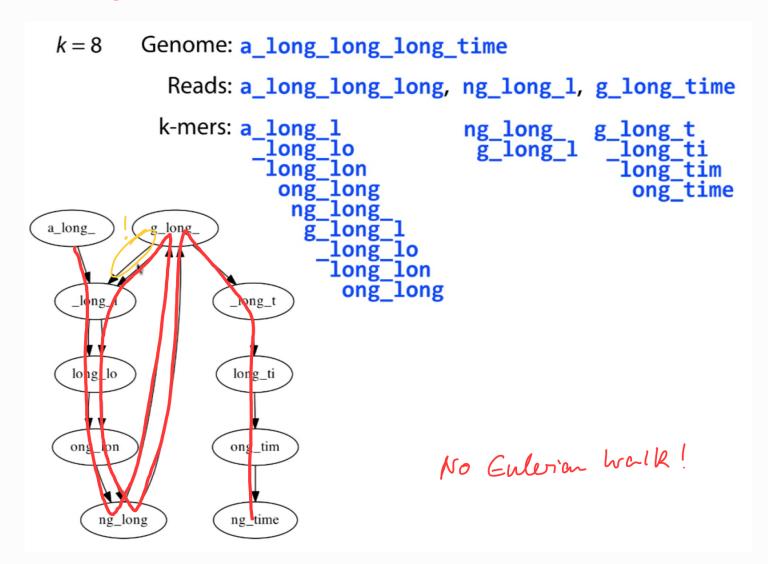
# a\_long\_long\_long\_time

Eulerian walk gives original genome!



Enterion Walk.

(tyrical K~ 30-50)



Overlap graphs and DeBruijn graphs can be used to represent "relationships" between substrings.

The provided algorithms can, in general, not solve the assembly problem in an "optimal way" but serve as useful heuristics.

There are more sophisticated methods out there that are often based on these type pf algorithms.

\*Medvedev & Pop What do Eulerian and Hamiltonian cycles have to do with genome assembly? PLoS Comput Biol. 2021

# **Classical problems in practice:**

- sequencing errors
- overlapping regions of fragments that are located on "far away" positions on DNA
- incomplete data (DNA not covered by resulting sequenced fragments)
- orientation of reads usually unknown
- repeats